



Aging & Memory Research Lab., School of Behavioral & Brain Sciences, Univ. of Texas at Dallas, Richardson, TX

### Introduction

Increased dietary fat intake leads to drastic alteration of systemic metabolism, obesity, insulin resistance, and type II diabetes, all potentially altering brain function and cognitive performance. Successful consolidation of many learning tasks reduces Ca<sup>2+</sup> dependent afterhyperpolarizations (AHPs) of hippocampal CA1 pyramidal neurons, increasing their intrinsic excitability. Since insulin has been shown to enhance memory retention, we investigated the effects of a chronic high-energy diet on systemic markers of metabolic dysfunction, on intrinsic excitability of CA1 hippocampal neurons, and on neural insulin sensitivity. Sex-dependent differences were also tested since estrogen effects both AHPs and insulin receptors.

## Methods

Subjects: Experiments were performed using behaviorally naive male & female Long Evans rats. Rats were bred use and maintained in our animal facility under conditions approved by the UT Dallas ACUC on a 12hr/12hr light/dark schedule with ad libitum access to food and water. Littermates were distributed across all treatment conditions to reduce variance, and males and females were socially housed.

**Diet:** Subjects in the Control diet cohort received a standard rat pellet diet with kcal energy contributions of: 20.7% protein, 35.8% fat, and 35% carbohydrate. Subjects in the HE pilot cohort received a diet of standard rat pellet augmented to achieve an energy contribution of: 15.6% protein, 57.6% fat, and 26.8% carbohydrate. Subjects received only their assigned diet from weaning (21 d) for approximately 15 weeks.

**Slice preparation:** Rats were anesthetized with isoflurane and decapitated. The brain was quickly hemisected and immersed in cooled sucrose-aCSF [in mM: 124 sucrose; 3 KCl; 1.3 MgSO<sub>4</sub>; 1.24 NaH<sub>2</sub>PO<sub>4</sub>; 2.4 CaCl<sub>2</sub>; 26 NaHCO<sub>3</sub>; 10 d-glucose]. After the brain chilled for 3-4 min, it was blocked and 400 µm slices cut using vibratomes then placed in room temperature (25°C) aCSF [in mM, 124 NaCl; 3 KCl; 1.3 MgSO<sub>4</sub>; 1.24 NaH<sub>2</sub>PO<sub>4</sub>; 2.4 CaCl<sub>2</sub>; 26 NaHCO<sub>3</sub>; 10 d-glucose]. Both aCSFs were continuously oxygenated (95%) O<sub>2</sub>: 5% CO<sub>2</sub>; ,pH 7.4).

**Current-clamp recordings:** Sharp electrodes were prepared from borosilicate glass filled with 3 M KCl (30-80 MΩ), and intracellular recordings made (using an AxoClamp-2B amplifier and National Instrument LabView interface) from submerged slices (31°C) . Measures of excitability included AHP peak amplitude, and duration, as well as measurements of AHP amplitude at varying intervals post-burst (to assess mAHP and sAHP components).

**Insulin Perfusion:** After baseline recordings, brain slices were perfused with the most effective dose of insulin (12.5 nM), as previously determined via dose response curve and recordings were repeated.

Glucose Tolerance Testing & Insulin Tolerance Testing (GTT/ITT): Rats were fasted for 8 hours and handled for 1 hour prior to testing. Basal plasma glucose levels (mg/dL) were assessed via glucometer immetiately prior to injecting either glucose (2g/kg)or insulin (1U/kg) or saline. Plasma glucose was then sampled every 15 min for 120min. **Systemic Measures:** Blood samples were collected during decapitation and plasma was frozen until use. Insulin was assessed using rat insulin ELISA kits (Abnova) and cortisol was assessed using CSCI ELISA kits (Abcam), using a ELx800 plate reader (BioTek) abd Gen5 software. Glycated hemoglobin levels were assessed with a DCA2000x analyzer (Bayar Laboratories) and DCA Systems Hemoglobin A1c test kits (Seimens).

# Sex-differences in Hippocampal CA1 Excitability and Systemic Responses in a High-Energy Diet Induced Prediabetic Rat Model E.L. Underwood & L.T. Thompson



(p < 0.05), much greater reductions were observed in HE diet neurons (p < 0.001)

